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SYNTHESIS AND ANTIFUNGAL ACTIVITY OF POLYPHENOL ETHER DERIVATIVES AGAINST PLANT PATHOGENIC FUNGI IN VITRO AND IN VIVO

Yue CUI,^{a,b} Hua-Guang LIU,^{a,b} Hua-Yang PAN,^{a,b} Shuang-Mei YAN,^{a,b} Ze-Xin QI,^{a,b} Xiao-Long ZHAO^{*a,b} and Du-Qiang LUO^{*a}

^aKey Laboratory of Medicinal Chemistry and Molecular Diagnosis of Ministry of Education; College of life science, Hebei University, Baoding 071002, P.R. China
^bCollege of Chemistry and Environmental Science, Hebei University, Baoding 071002, P.R. China

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Phytopathogenic fungi have been considered as an enormous threat in the agricultural system. In our search of new antifungal natural products, twenty-seven polyphenol ether derivatives were synthesized. Their structures were elucidated based on extensive spectroscopic analysis. The results suggest that compound **8**, **13**, **15** and **21** exhibited strong antifungal activities against *R. solani* and *S. sclerotiorum in vitro*, and compound **13** displayed the best antifungal efficacy toward *E.* graminis *in vivo*. This work provides an effective strategy for searching antifungal candidate agents.



INTRODUCTION

Polyphenols are secondary metabolites of plats and widely exist in nature. They are a class of compounds with a wide range of pharmacological properties. At present, studies on polyphenols mainly focus on their bioavailability, antioxidant and anti-cancer activity, and bacteriostasis.¹ For example, considerable recent interest has concentrated on bioactive phenolic compounds in grape, as they possess many biological activities, such as antioxidant, cardioprotective, anti-cancer, anti-inflammation, anti-ageing and antimicrobial properties.² It is reported that berry phenolics have extensive beneficial effects because of their antioxidant and anti-inflammatory properties.³ Y. Zhang found that Chroogomphus rutilus polyphenol extracts revealed a higher antioxidant, anti-inflammatory, and cytotoxic activities.⁴ L. Delgado-Roche found that polyphenols have certain cytotoxicity to colorectal cancer cells, which can inhibit tumor growth and induce apoptosis.⁵ In terms of bacteriostasis, research has shown that polyphenols from Amygdalus

^{*} Correspondence author : duqiangluo999@126.com (Du-Qiang Luo); longlong_666@sina.com (Xiao-Long Zhao)

pcdunculata Pall seed coat had good antibacterial activity against *Escherichia coli*, *Staphylococcus aureus*, *Bacillus cereus*, and *Bacillus subtilis*.⁶ Simultaneously, polyphenols were proved to play a beneficial role in the prevention and development of chronic diseases related to inflammation, such as diabetes, obesity, neurodegeneration, and cardiovascular diseases.⁷

Gallic acid (3,4,5 trihydroxybenzoic acid), a phenolic acid, is known to form various derivatives having a variety of the aforementioned biological functions.8 D. Wang reported that gallic acid inhibited the activation of Epidermal Growth Factor Receptor (EGFR), repressed the proliferation and elevated apoptosis of Nonsmallcell Lung Cancer (NSCLC) cells, and it has been confirmed in animal experiments that gallic acid exhibited an inhibitory effect on tumor growth in vivo.9 Research has shown that gallic acid interacts with paclitaxel to increase its cytotoxic effect and cell induction.¹⁰ According to research focused on gallic acid, it was found that the gallic acid can induce apoptosis of human gastric cancer cells.¹¹ Studies also suggested that gallic acid is antiinflammatory via attenuating Lipopolysaccharide (LPS)-induced neuroinflammation, oxidative stress, and protein conjugation.¹²

multitude Although a of galloylated polyphenolic compounds distribute in nature, gallovlated phenols also need to be produced synthetically to influence their biological properties.¹³ In this paper, a series of alkyl benzoate ether derivates were synthesized, and the antifungal activity of the prepared 27 compounds against plant pathogenic fungi in vitro and in vivo were investigated.

RESULTS AND DISCUSSION

In vitro activity against phytopathogenic fungi

The antifungal activity of the synthesized polyphenol ether derivatives against *Rhizoctonia solani* and *Sclerotinia sclerotiorum* were tested, and the results were listed in Table 1. It is clear that among this series of prepared compounds, compound **8**, **13**, **15** and **21** exhibited greater antifungal activity in comparison with other

compounds, with the mycelial growth inhibition values of 60.7 ± 3.3 (8), 66.2 ± 1.7 (13), 73.7 ± 3.3 (15), 64.9 ± 5.6 (21) for *R. solani*, and 73.0 ± 3.9 (8), 68.9 ± 0.5 (13), 73.3 ± 4.2 (15), 63.9 ± 6.2 (21) for *S. sclerotiorum*, respectively. Particularly, compound 15 was the most potent that displayed the most efficient mycelial growth inhibition action toward *R. solani* and *S. sclerotiorum*.

To further investigate the antifungal effect of compound 8, 13, 15 and 21, other seven pathogenic fungi including Alternaria Alternate, Botrytis Cinerea, Fusarium Graminearum, Fusarium Oxysporium, Glomerella Cinglata, Gaeumannomyces Gramims, Rhizoctonia Solani, Sclerotinia Sclerotiorum, and Venturia Nashicola were selected. The inhibition activity against these fungi as well as R. solani and S. sclerotiorum of compound 8, 13, 15 and 21 were given in Fig. 1. The results showed that these compounds were more susceptible to R. solani and S. sclerotiorum comparing with other fungi. Furthermore, it was noted that besides R. solani and S. sclerotiorum, compound 8 also exhibited notable antifungal activity against B. Cinerea and F. Oxysporium with the inhibition rates of mycelial growth of 48.8% and 50.0%, and compound 21 also displayed prominent antifungal action toward A. Alternate, B. Cinerea and F. Oxysporium with the inhibition rates of mycelial growth of 54.8%, 63.5% and 54.4%, respectively.

In vivo activity against Erysiphe graminis

The control effects on wheat powdery mildew of the series of polyphenol ether derivatives in greenhouse were also tested. Their fungicidal activities against E. graminis at concentration of 500 µg/ml were determined in vivo, and the results are listed in Table 2. The results showed that had strong activity against compound 13 E. graminis in vivo with the preventive and curative effects of 66.9 and 72.1%, respectively, although the effects are weaker than those of standard fungicide Triadifeon. However. compound 8, 15 and 21 that possessed outstanding antifungal effect in vitro had almost no antifungal activity toward E. graminis in green house.

| | Inhibition of mycelial growth (%) (±SD) ^a | | |
|-----------------|--|-----------------|--|
| No. of compound | R. solani | S. sclerotiorum | |
| 1 | (14.6±3.9) k | (8.4±2.1) K | |
| 2 | (26.7±6.9) fg | (7.3±1.9) K | |
| 3 | (16.0±3.5) ji | (19.8±1.4) JI | |
| 4 | (45.3±3.4) d | (35.7±9.2) FGH | |
| 5 | (44.3±8.7) d | (25.8±1.6) HI | |
| 6 | (31.5±7.5) ef | (40.4±2.4) FG | |
| 7 | (14.3±3.8) k | 0 L | |
| 8 | (60.7±3.3) bc | (73.0±3.9) A | |
| 9 | (22.3±6.0) hgi | (34.0±4.6) FG | |
| 10 | (23.6±3.6) gh | (41.1±2.2) FE | |
| 11 | (37.1±2.4) e | (47.4±1.8) DE | |
| 12 | (28.3±3.0) fg | (49.3±4.2) DC | |
| 13 | (66.2±1.7) b | (68.9±0.5) AB | |
| 14 | (10.8±3.4) k | (17.6±3.7) J | |
| 15 | (73.7±3.3) a | (73.3±4.2) A | |
| 16 | 01 | (35.5±4.0) FGH | |
| 17 | (17.9±0.5) hji | (22.1±1.7) JI | |
| 18 | (56.9±2.8) c | (55.3±7.6) C | |
| 19 | (17.1±1.6) hji | (38.7±2.7) FG | |
| 20 | (32.6±3.1) ef | (38.0±5.1) FG | |
| 21 | (64.9±5.6) b | (63.9±6.2) B | |
| 22 | 01 | 0 L | |
| 23 | 01 | (3.6±1.3) KL | |
| 24 | (18.6±0.7) hji | (7.1±2.2) K | |
| 25 | (17.6±1.0) ji | (9.2±0.8) K | |
| 26 | (27.4±4.3) fg | (29.1±4.1) HG | |
| 27 | (37.0±4.6) e | (33.5±4.5) FGH | |



Fungi toxicity of the prepared compounds on mycelial growth inhibition of R, solari and S, sclerotiorum



Fig. 1 – Antifungal activity spectra of compound **8**, **13**, **15** and **21** against nine pathogenic fungi. Error bars represent the standard error of the mean of three replicates.

| No. of compound | Dose (mg/L) – | Protective effects (%) (±SD) a | Curative effects (%) (±SD) b |
|-----------------|---------------|--------------------------------|------------------------------|
| | | 8 days | 8 days |
| 1 | 500 | (15.0±3.3) jk | (12.7±2.8) KJ |
| 2 | 500 | 01 | (18.5±4.7) KJI |
| 3 | 500 | 01 | (38.4±6.0) DE |
| 4 | 500 | (20.9±1.3) hjki | (39.0±4.8) DE |
| 5 | 500 | (37.8±6.3) cde | (28.8±3.1) HFG |
| 6 | 500 | (13.5±5.8) k | (33.3±3.0) EDF |
| 7 | 500 | (17.0±5.3) jki | (11.0±3.0) K |
| 8 | 500 | (37.3±8.2) cde | (40.0±1.0) D |
| 9 | 500 | (25.4±6.2) hfg | (27.6±5.1) HFG |
| 10 | 500 | (32.5±6.7) cdef | 0 L |
| 11 | 500 | 01 | (13.8±2.8) KJ |
| 12 | 500 | (35.5±1.2) cde | 0 L |
| 13 | 500 | (66.9±4.3) b | (72.1±6.8) B |
| 14 | 500 | (40.0±2.4) c | (34.3±6.6) EDF |
| 15 | 500 | (31.4±7.6) cdef | (47.5 ±2.2) C |
| 16 | 500 | (29.8±1.9) efg | (14.1±6.1) KJ |
| 17 | 500 | 01 | (36.1±2.1) EDF |
| 18 | 500 | (30.9±6.6) def | (32.5±6.4) EDF |
| 19 | 500 | (14.7±1.6) jk | (49.2±7.7) C |
| 20 | 500 | (39.2±3.4) cd | (21.1±6.7) HJI |
| 21 | 500 | (22.3±4.8) hgji | (31.0±0.5) EGF |
| 22 | 500 | (15.1±3.0) jk | (30.5±5.2) EGF |
| 23 | 500 | (13.8±3.7) jk | (37.7±5.9) EDF |
| 24 | 500 | (3.9±4.2)1 | (16.0±4.5) KJI |
| 25 | 500 | (24.7±8.0) hfgi | (14.7±3.6) KJI |
| 26 | 500 | (13.6±4.2) k | (19.1±5.8) KJI |
| 27 | 500 | (32.8±5.5) cdef | (23.0±3.8) HGI |
| Triadifeon | 250 | (82.58±1.95) a | (88.3±1.7) A |

Table 2

In vivo control of Erysiphe graminis with protective and curative spray applications of polyphenol ether derivatives

EXPERIMENTAL SECTION

Materials and Measurements

Alkanols were purchased from Shanghai Chemical Factory (China). Gallic acid, Dicyclohexylcarbodiimide (DCC), trans,trans-farnesyl bromide, 4-bromo-2-methyl-2-butene, 3,5-dihydroxybenzoic acid and chloromethyl methyl ether were purchased from Aldrich Chemical Co. (Milwaukee, WI). The organic solvents were purified and dried using appropriate procedures. EI-MS spectra were measured with a VG Autospec3000 mass spectrometer (VG, England). ¹H, ¹³C-NMR spectra were recorded on Bruker AM-400 and DRX-500 spectrometer (Karlsruhe, Germany). The chemical shifts were reported in ppm (δ) relative to the internal standard tetramethylsilane (TMS). Air- and/or moisture-sensitive reactions were carried out under an argon or nitrogen atmosphere.

Synthesis

Synthesis of alkyl gallates ester

To a solution of gallic acid (2.00 mM) and the corresponding alcohol (CH3CH2OH, (CH3)2CHCH2CH2OH, CH₃(CH₂)₁₆CH₂OH, CH₃OH, (CH₃)₂CHOH, (CH₃)₂CHCH₂CH₂OH, CH₃CH₂CH₂(CH₃)CHOH, 2.00 mM) in Tetrahydrofuran (THF, 10 mL) cooled at 0°C was added a solution of DCC (4.2 mM) in THF (10 mL). After stirring for 10 h, the solvent of the resulted mixture was removed under reduced pressure. The residue was extracted with ethyl acetate five times and filtered. The filter was washed successively with 4 M HCl solution, saturated NaHCO3 solution, and water, and then dried over Na₂SO₄ and evaporated. The crude products were purified by column chromatography on silica gel with petroleum ether/ethyl acetate (4:1, v/v) as eluent. The synthetic route of the esters was shown in Scheme 2.

Synthesis of 3,5-dihyrdroxybenzaldehyde



Scheme 2 – Synthesis of polyphenol ether derivatives.

In order to synthesize polyphenol ether derivatives, anhydrous K_2CO_3 (2 mM), anhydrous acetone (10 mL) and R-X (trans, trans-farnesyl bromide, 4-bromo-2-methyl-2-butene, chloromethyl methyl ether) were added to the solution of corresponding alkyl gallates ester (1 mM) or 3,5dihydroxybenzaldehyde (1 mM). Then the reaction mixture was heated under reflux for 24h. The solvent was removed under reduced pressure. The residue was extracted with ethyl acetate several times and filtered. The filter was washed successively with 4M HCl solution, saturated NaHCO₃ solution and water, and then dried over Na₂SO₄ and evaporated. The crude products were purified by column chromatography on silica gel with petroleum ether/ethyl acetate (7:1, v/v) as eluent. Structures of the synthesized esters were illustrated in Table 3.

Characterization of the prepared compounds

Ethyl 4-((2E,6E),2,6,11-trimethyldodeca-2,6,10trienyloxy)-3,5-dihydroxybenzoate (C₂₄H₃₄O₅, 1): obtained in 34% yield as yellow oil. EI-MS: m/z 402M⁺ (4), 265 (3), 249 (12), 219 (13), 205 (26), 198 (55), 183 (19), 170 (30), 153 (64), 149 (34), 137 (51), 121 (43), 107 (33), 95 (48), 81(100), 69 (84), 55 (18); ¹³C NMR (CDCl₃, 400 MHz) δ : 166.3 (s, CO), 149.1 (s), 145.3 (s), 137.3 (s), 135.7 (s), 131.4 (s), 126.6 (s), 124.3 (d), 123.4 (d), 118.6 (d), 109.5 (d), 69.9 (t, OCH2), 61.1 (t, OCH₂), 39.6 (t), 32.0 (t), 26.7 (t), 26.1 (t), 25.7 (q, CH₃), 17.7 (q, CH₃), 16.4 (q, CH₃), 16.0 (q, CH₃), 14.7 (q, CH₃), 14.2 (q, CH₃); ¹H NMR (CDCl₃, 400 M) δ : 7.23 (s, 2H, ArH), 4.63 (d, J=6.0 Hz, 2H, OCH₂), 4.34 (dd, J=5.7 Hz, J=11.4 Hz, 2H, OCH₂), 5.73 (s, 2H, OH), 5.54 (t, 1H, CH), 5.08 (t, 2H, CH), 2.06 (m, 2H, CH₂), 1.97 (m, 2H, CH₂), 1.26 (t, CH₃), 1.60 (s, 6H, CH₃), 1.68 (s, 3H, CH₃), 1.71 (s, 3H, CH₃).

Ethyl 4-(3-methylbut-2-enyloxy)-3,5-dihydroxybenzoate (C₁₄H₁₈O₅, 2): obtained in 45% yield as a colorless solid. Positive FAB-MS M+H⁺ 267; ¹³C NMR (CDCl₃, 400 MHz) δ : 166.3 (s, CO), 149.1 (s), 149.1 (s), 141.7 (s), 137.8 (s), 126.5 (s), 119.1 (d), 109.5 (d), 109.5 (d), 70.0 (t), 61.1 (t), 25.8 (q), 18.0 (q), 14.2 (q); ¹H NMR (CDCl₃, 400 M) δ : 7.25 (s, 2H, ArH), 5.85 (s, 2H, OH), 5.23 (t, 1H, CH), 4.62 (d, J=6.0 Hz, 2H, OCH₂), 4.34 (dd, J=5.7 Hz, J=11.4 Hz, 2H, OCH₂), 1.77 (s, 3H, CH₃), 1.66 (s, 3H, CH₃), 1.37 (s, 3H, CH₃).

Ethyl 3-(3-methylbut-2-enyloxy)-4,5-dihydroxybenzoate (C₁₄H₁₈O₅, 3): obtained in 56% yield as yellow colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 166.2 (s, CO), 145.3 (s), 143.0 (s), 139.1 (s), 136.8 (s), 121.6 (s), 118.5 (d), 110.4 (d), 65.8 (t, OCH₂), 60.8 (t, OCH₂), 25.5 (q), 17.9 (q), 14.0 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.39 (s, 1H, ArH), 7.24 (s, 1H, ArH), 4.59 (d, J=6.8 Hz, H, OCH₂), 4.31 (dd, J=7.1 Hz, J=15.1 Hz, 2H, OCH₂), 5.45 (t, 1H, CH), 1.40 (t, 3H, CH₃), 1.77 (s, 3H, CH₃), 1.93 (s, 3H, CH₃).

Isopentyl 4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-3,5-dihydroxybenzoate (C₂₇H₄₀O₅,4): obtained in 45% yield as colorless oil. EI-MS: m/z 444 M⁺ (5), 375 (4), 362 (4), 331 (3), 291 (18), 240 (4), 205 (15), 191 (25), 183 (40), 170 (86), 153 (57), 137 (41), 121 (37), 93 (52), 81 (83), 69 (100), 55 (22); HR-ESI M+Na⁺ 467.2772, calc. 467.2773; ¹³C NMR (CDCl₃, 400 MHz) δ : 166.9 (s, CO), 149.3 (s), 149.3 (s), 144.6 (s), 137.5 (s), 135.5 (s), 131.2 (s), 125.8 (s), 124.2 (d), 123.4 (d), 118.8 (d), 109.4 (d), 109.4 (d), 25.6 (d), 69.6 (t, OCH₂), 63.9 (t, OCH₂), 39.6 (t), 39.5 (t), 37.2 (t), 26.6 (t), 26.1 (t), 25.0 (q), 22.4 (q), 17.6 (q), 16.4 (q), 16.3 (q), 15.9 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.29 (s, 2H, ArH), 4.32 (t, 2H, OCH₂), 4.69 (d, J=7.3 Hz, 2H, OCH₂), 5.07 (br 2H, CH), 5.52 (t, 1H, CH), 6.54 (br, 2H, OH), 097 (d, J=12.9 Hz, 6H, CH₃), 1.59 (br s, 6H, CH₃), 1.67 (s, 3H, CH₃), 1.75 (s, 3H, CH₃), 1.95 (m, 1H, CH), 2.07 (m, 8H, CH₂).

Isobutyl 4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-3,5-dihydroxybenzoate) (C₂₆H₃₈O₅, 5): obtained in 32% yield as yellow oil. HR-ESI M+Na⁺ 453.2617, calc. 453.2616; ¹³C NMR (CDCl₃, 400 MHz) δ : 166.7 (s, CO), 149.3 (s), 144.6 (s), 137.7 (s), 135.6 (s), 131.2 (s), 126.2 (s), 124.4 (d), 123.5 (d), 123.5 (d), 119.0 (d), 109.6 (d), 109.6 (d), 26.7 (d), 71.3 (t, OCH₂), 69.8 (t, OCH₂), 39.6 (t), 39.6 (t), 26.7 (t), 26.2 (t), 25.5 (q, CH₃), 19.1 (q, CH₃), 19.1 (q, CH₃), 17.6 (q, CH₃), 16.4 (q, CH₃), 15.9 (q, CH₃); ¹H NMR (CDCl₃, 400 M) δ : 6.22 (s, 2H, ArH), 4.68 (d, J=7.4 Hz, 2H, OCH₂), 4.08 (d, J=6.5 Hz, 2H, OCH₂), 5.53 (t, 1H, CH), 5.08 (t, 2H, CH), 2.01 (m, 1H, CH), 0.97 (d, J=18.2 Hz, 6H, CH₃), 1.67 (s, 6H, CH₃), 1.60 (s, 6H, CH₃), 2.08 (m, 8H, CH₂).

Isobutyl 3-((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-4,5-dihydroxybenzoate (C26H38O5, 6): obtained in 22% yield as yellow oil. Positive FAB-MS M+H+ 431; HRESI-MS M+Na⁺ 453.2629, cacl. 453.2616; ¹³C NMR (CDCl₃, 400 MHz) *δ*: 166.8 (s, CO), 150.0 (s), 143.6 (s), 143.0 (s), 137.3 (s), 135.9 (s), 131.7 (s), 122.4 (s), 124.6 (d), 123.8 (d), 118.9 (d), 111.0 (d), 106.4 (d), 28.2 (d), 71.3 (t, OCH₂), 66.5 (t, OCH2), 40.0 (t), 39.8 (t), 27.0 (t), 26.5 (t), 26.0 (q, CH3), 19.5 (q, CH3), 19.5 (q, CH3), 18.0 (q, CH3), 17.0 (q, CH₃), 16.3 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ: 7.34 (s, 1H, ArH), 7.22 (s, 1H, ArH), 4.65 (d, J=5.3 Hz, 2H, OCH₂), 4.07 (d, J=5.3 Hz, 2H, OCH2), 5.07 (br s, 2H, OH), 5.89 (br t, 1H, CH), 5.44 (br t, 2H, CH), 2.04 (br t, 8H, CH₂), 0.98 (d, J=6.7 Hz, 6H, CH₃), 1.58 (br s, 6H, CH₃), 1.67 (d, J=12.7 Hz, 3H, CH₃), 1.76 (d, J=21.8 Hz, 3H, CH₃).

Octadecyl 4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-3,5-dihydroxybenzoate (C₄₀H₆₆O₅, 7): obtained in 65% as colorless oil. HR-ESI M+Na⁺ 649.4816, calc. 649.4807; ¹³C NMR (CDCl₃, 400 MHz) δ : 166.8 (s, CO), 149.3 (s), 149.3 (s),144.8 (s), 137.4 (s), 135.6 (s), 131.3 (s), 126.1 (s), 124.2 (d), 123.4 (d), 118.8 (d), 109.5 (d), 109.5 (d), 69.7 (t, OCH₂), 65.5 (t, OCH₂), 39.6 (t), 39.6 (t), 39.6 (t), 31.1-29.3 (br t,11CH₂), 29.3 (t), 28.6 (t), 26.7 (t), 26.2 (t), 26.0 (t), 25.7 (q), 22.7 (q), 16.4 (q), 16.0 (q), 14.1 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.27 (s, 2H, ArH), 6.29 (br s, 2H, OH), 4.28 (t, 2H, OCH₂), 4.68 (d, J=7.5 Hz, 2H, OCH₂), 5.26 (t, 1H, CH), 5.08 (t, 2H, CH), 0.88 (t, 3H, CH₃), (br s, 6H, CH₃), 1.77 (s, 3H, CH₃), 1.75 (s, 3H, CH₃), 2.07 (m, 8H, CH₂), 1.26 (br m, 32H, CH₂).

Methyl 3,4-bis(3-methylbut-2-enyloxy)-5-hydroxybenzoate (C₁₈H₂₄O₅, 8): obtained in 65% as colorless solid. ¹³C NMR (CDCl₃, 400 MHz) δ: 166.8 (s, CO), 151.3 (s), 149.6 (s), 140.4 (s), 138.5 (s), 138.3 (s), 125.4 (s), 119.4 (d), 119.2 (d), 109.4 (d), 106.8 (d), 69.4 (t, OCH₂), 66.2 (t,OCH₂), 65.7 (t,OCH₂), 52.1 (q, OCH₃), 25.8 (q, CH₃), 25.8 (q, CH₃), 18.2 (q), 17.9 (q); ¹H NMR (CDCl₃, 400 MHz) δ: 7.28 (s, 1H, ArH), 7.20 (s, 1H, ArH), 3.88 (s, OCH₃), 4.67 (d, J=7.5 Hz, 2H, OCH₂), 4.60 (d, J=5.4 Hz, 2H, OCH₂), 5.86 (s, 1H, OH), 5.50 (br t, 2H, CH), 1.66 (s, 3H, CH₃), 1.75 (br s, 6H, CH₃), 1.80 (s, 3H, CH₃).

Methyl3,4,5-tris(3-methylbut-2-enyloxy)benzoate($C_{23}H_{32}O_5$, 9): obtained in 56% yield as colorless oil. HR-ESI $M+1^+$ 389.2327, calc. 389.2344; 13 C NMR (CDCl₃, 400 MHz) δ : 167.2 (s, CO), 149.3 (s), 145.8 (s), 143.6 (s), 141.2 (s),139.3 (s), 137.5 (s), 121.4 (s), 119.2 (d), 118.8 (s), 110.9 (d),118.8 (d), 110.9 (d), 109.5 (d), 106.2 (d), 69.7 (t, OCH₂), 60.5 (t, OCH₂), 66.1 (t, OCH₂), 52.1 (q, OCH₃), 25.8 (q), 25.7 (q),25.5 (q), 21.0 (q), 18.1 (q), 17.9 (q); 1 H NMR (CDCl₃, 400 MHz) δ : 7.34 (s, 1H, ArH), 7.23 (s, 1H, ArH), 4.65 (d)

J=7.5 Hz, 2H, OCH₂), 4.59 (d, J=6.6 Hz, 4H, OCH₂), 4.13 (br s, 3H, 3H), 3.88 (s, 3H, OCH₃), 1.64 (s, 6H,CH₃), 1.75 (br s, 6H, CH₃), 2.05 (s,6H, CH₃).

Methyl 3,4-bis((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-5-hydroxybenzoate (C38H56O5, 10): obtained in 65% yield as colorless oil. HR-ESI M+Na⁺ 615.4025, cacl. 615.4025; ¹³C NMR (CDCl₃, 400 MHz) δ: 166.7 (s, CO), 151.3 (s), 149.6 (s), 143.7 (s), 141.4 (s), 138.3 (s), 135.3 (s), 135.3 (s), 131.1 (s), 131.1 (s), 125.3 (s), 124.2 (d), 124.2 (d), 123.5 (d), 123.5 (d), 119.1 (d), 119.0 (d), 109.4 (d), 106.5 (d), 69.1 (t, OCH₂), 65.6 (t, OCH₂), 39.6 (t), 39.6 (t), 39.5 (t), 39.5 (t), 26.6 (t), 26.6 (t), 26.1 (t), 26.1 (t), 52.0 (q, OCH₃), 25.8 (q, CH3), 25.6 (q, CH3), 17.6 (q, CH3), 17.6 (q, CH3), 16.6 (q, CH₃), 16.2 (q, CH₃), 15.9 (q, CH₃), 15.9 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ: 7.23 (s, 1H, ArH), 7.16 (s, 1H, ArH), 3.83 (s, 3H, OCH₃), 4.65 (d, J=7.4 Hz, OCH₂), 4.58 (d, J=6.5 Hz, OCH₂), 5.99 (s, 1H, OH), 1.54 (s, 6H, CH₃), 1.62 (s, 6H, CH₃), 1.66 (s, 6H, CH₃), 1.75 (s, 6H, CH₃); 5.48 (br t, 2H, CH), 5.05 (m, 4H, CH), 2.06 (br, m, 8H, CH₂), 1.94 (br t, 8H, CH2).

Methyl 4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-3,5-dihydroxybenzoate (C₂₃H₃₂O₅, 11): obtained in 67% yield as colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 166.8 (s, CO), 149.3 (s), 144.8 (s), 137.4 (s), 135.6 (s), 131.3 (s), 126.0 (s), 124.3 (d), 123.4 (d), 118.8 (s), 109.4 (s), 69.6 (t, OCH₂), 61.3 (t, OCH₂), 39.6 (t), 39.6 (t), 26.6 (t), 26.1 (t), 25.6 (q, CH₃), 17.6 (q, CH₃), 16.4 (q, CH₃), 15.9 (q, CH₃), 14.2 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ : 6.38 (s, 2H, ArH), 4.68 (d, J=7.5 Hz, 2H, OCH₂), 4.34 (dd, J=7.1 Hz, J=14.2 Hz, 2H, OCH₂), 5.52 (t, 1H, CH), 5.07 ((br t, 2H, CH), 1.37 (t, 3H, CH₃), 1.59 (br s, 6H, CH₃), 1.65 (s, CH₃), 1.71 (s, CH₃), 2.04 (br tm, 8H, CH₂).

3,5-bis(3-methylbut-2-enyloxy)benzaldehyde (C₁₇H₂₂O₃, **12):** obtained in yield 42% as colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 191.9 (d, CHO), 160.3 (s, CO), 160.3 (s, CO), 138.7 (s), 138.2 (s), 138.2 (s), 118.9 (d), 118.9 (d), 108.3 (d), 107.7 (d), 107.7 (d), 65.1 (t, OCH₂), 65.1 (t, OCH₂), 25.7 (q, CH₃), 25.7 (q, CH₃), 18.1 (q, CH₃), 18.1 (q, CH₃); ¹H NMR (CDCl₃, 400 M) δ : 9.97 (s, CHO), 7.11 (s, 2H, ArH), 6.84 (s, 1H, ArH), 4.64 (d, J=6.7 Hz, 4H, OCH₂), 5.59 (br t, 2H, CH), 1.93 (br s, 6H, CH₃), 1.86 (br s, 6H, CH₃).

Isopropyl 3,4-bis(3-methylbut-2-enyloxy)-5-hydroxybenzoate (C₂₀H₂₈O₅, 13): obtained in 45% as colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 165.8 (s, CO), 151.2 (s), 149.5 (s), 140.2 (s), 138.3 (s), 138.1 (s), 126.1 (s), 119.6 (d), 119.4 (d), 109.3 (d), 106.6 (d), 68.3 (d), 69.3 (t, OCH₂), 65.6 (t, OCH₂), 25.7 (q), 25.7 (q), 21.9 (q), 21.9 (q), 18.2 (q), 17.8 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.63 (s, 1H, ArH), 7.55 (s, 1H, ArH), 5.02 (d=7.5 Hz, 2H, OCH₂), 4.96 (d, J=6.7 Hz, 2H, OCH₂), 5.56 (m, 1H, CH), 5.83 (br t, 2H,CH), 6.32 (s, 1H, OH), 1.70 (d, J=6.5 Hz, 6H, CH₃), 2.00 (s, 3H, CH₃), 2.01 (s, 3H, CH₃), 2.05 (s, 3H, CH₃), 2.15 (s, 3H, CH₃).

Isopropyl 3,4,5-tris(3-methylbut-2-enyloxy)benzoate (C2sH36Os, 14): obtained in 46% yield as colorless oil. HR-ESI M+Na⁺ 439.2457, calc. 439.2460; ¹³C NMR (CDCl₃, 400 MHz) δ : 165.9 (s, CO), 152.6 (s), 152.6 (s), 142.0 (s), 138.3 (s), 137.6 (s), 125.4 (s), 123.4 (s), 121.6 (d), 119.9 (d), 108.4 (d), 108.4 (d), 69.2 (t, OCH₂), 68.3 (d, OCH₂), 66.0 (t, OCH₂), 66.0 (t, OCH₂), 25.5 (q), 22.0 (q), 18.2 (q), 17.9 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.26 (s, 2H, ArH), 4.61 (d, J=6.6 Hz, 4H, OCH₂), 4.56 (d, J=7.4 Hz, 2H, OCH₂), 5.23 (m, 1H, CH), 5.54 (br t, 3H, CH), 1.36 (d, J=6.0 Hz, 6H, CH₃), 1.54 (s, 3H, CH₃), 1.66 (s, 3H, CH₃), 1.77 (br s, 6H, CH₃).

Isopropyl 4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10trienyloxy)-3,5-dihydroxybenzoate (C₂₅H₃₆O₅, 15): obtained in 36% as colorless solid.¹³C NMR (CDCl₃, 400 MHz) δ: 166.2 (s, CO), 149.5 (s), 145.3 (s), 137.7 (s), 136.0 (s), 131.6 (s), 127.2 (s), 124.6 (d), 123.8 (d), 119.2 (d), 109.8 (d), 109.8 (d), 68.9 (d, CH), 70.2 (t, OCH₂), 40.0 (t), 40.0 (t), 27.1 (t), 26.5 (t), 26.0 (q, CH₃), 22.2 (q, CH₃), 22.2 (q, CH₃), 18.0 (q, CH₃), 16.8 (q, CH₃), 16.3 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ : 7.23 (s, 2H, ArH), 4.66 (d, J=7.5 Hz, 2H, OCH₂), 5.21 (m, 1H, CH), 5.54(t, 1H, CH), 5.21 (br t 2H, CH), 5.98 (s, 2H, OH), 1.34 (br s, 6H, CH₃), 1.68 (br s, 6H, CH₃), 1.76 (br s, 6H, CH₃), 1.98 (br t, 4H, CH₂), 2.00 (br m, 4H, CH₂).

3,5-bis((2E,6E)-**3,7,11-trimethyldodeca-2,6,10***trienyloxy)benzaldehyde* (C₃₇H₅₄O₃,16): obtained in 43% yield as yellow oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 192.0 (s, CHO), 160.5 (s), 141.8 (s), 141.8 (s), 138.3 (s), 135.5 (s), 135.5 (s), 135.5 (s), 131.3 (s), 131.3 (s), 124.3 (d), 123.6 (d), 123.6 (d), 118.9 (d), 118.9 (d), 108.4 (d), 107.9 (d), 107.9 (d), 65.8 (OCH₂), 65.3 (OCH₂), 65.3 (OCH₂), 39.7 (t), 39.7 (t), 39.6 (t), 26.7 (t), 26.7 (t), 26.2 (t), 26.2 (t), 25.7 (q), 25.7 (q), 17.7 (q), 17.7 (q), 16.7 (q), 16.7 (q), 16.0 (q), 16.0 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 9.89 (s, 1H, CHO), 7.02 (s, 2H, ArH), 6.74 (s, 1H, ArH), 4.56 (d, J=6.5 Hz, 4H, OCH₂), 5.49 (t, 2H, CH₃), 1.96 (br s, 6H, CH₃), 2.15-2.04 (br s, 16H, CH₂).

3-((2E,6E)-3,7,11-trimethyldodeca-2,6,10-trienyloxy)-5hydroxybenzaldehyde (C₂₂H₃₀O₃, 17): obtained in 34% yield as colorless solid. ¹³C NMR (CDCl₃, 400 MHz) δ : 191.7 (s, CHO), 160.3 (s), 157.4 (s), 141.9 (s), 138.7 (s), 138.7 (s), 135. 9 (s), 131.3 (s), 124.4 (d), 123.7 (d), 119.0 (d), 108.9 (d), 108.3 (d), 65.5 (t, OCH₂), 39.7 (t), 39.6 (t), 26.8 (t), 26.3 (t), 26.8 (t), 25.6 (q, CH₃), 17.6 (q, CH₃), 16.7 (q, CH₃), 16.0 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ : 9.87 (s, CHO), 7.00 (s, 1H, ArH), 6.67 (s, 1H, ArH), 5.47 (t, 1H, CH), 5.10 (br t, 3H, 2H), 4.57 (d, J=6.5 Hz, 2H, OCH₂), 1.59 (br s, 6H, CH₃), 1.74 (s, 3H, CH₃), 1.96 (s, 3H, CH₃), 2.13-2.03 (br t, 8H, CH₂).

Isopentyl 4-(3-methylbut-2-enyloxy)-3,5-dihydroxybenzoate (C₁₇H₂₄O₅, 18): obtained in 45% yield as white solid. ¹³C NMR (CDCl₃, 400 MHz) δ: 166.8 (s, CO), 149.2 (s), 149.2 (s), 141.5 (s), 137.4 (s), 126.1 (s), 119.1 (d), 109.4 (d), 109.4 (d), 25.0 (d), 69.8 (d, OCH₂), 63.9 (t, OCH₂), 37.2 (t), 26.0 (q), 25.8 (q), 22.4 (q), 22.4 (q); ¹H NMR (CDCl₃, 400 MHz) δ: 7.23 (s, 2H, ArH), 6.24 (br 2H, OH), 5.48 (t, 1H, CH), 4.28 (t, 2H, OCH₂), 0.92 (d, J=6.6 Hz, 6H, CH₃), 1.61 (s, 6H, CH₃), 1.77 (br, 3H, CH₂, CH).

Pentan-2-yl3,4-bis(3-methylbut-2-enyloxy)-5-hy-
droxybenzoatedroxybenzoate(C22H32O5, 19):obtainedin45%yieldscolorlessoil.13CNMR(CDCl3, 400 MHz) δ :166.9(s, CO),151.2(s),149.4(s),140.3(s),138.2(s),138.1(s),126.1(s),19.5(d),119.4(d),109.2(d),106.5(d),71.7(d),69.3(t,OCH2),65.6(t,OCH2),35.7(t),27.5(t),25.7(q),22.5(q),20.0(q),18.2(q),14.0(q);14NMRCDCl3,400MHz) δ :7.24(s,1H,ArH),7.16(s,1H,ArH),4.57(d,12.4(br,13.4(br,14.5(br,14.5(br,14.6(c),14.6(c),14.7(br,14.8(c),14.9(c),14.9(c),14.9(c),14.9(c),14.9(c),14.9(c),14.9(c),14.9(c),14.9(c),14.9(c),<t

Pentan-2-yl 4-(3-methylbut-2-enyloxy)-3,5-dihydroxybenzoate (C₁₇H₂₄O₅, **20**): obtained in 21% yield as a white solid. ¹³C NMR (CDCl₃, 400 MHz) δ : 166.1 (s, CO), 149.1 (s), 141.6 (s), 137.3 (s), 126.8 (s), 123.3 (s), 119.1 (d), 109.4 (d), 109.4 (d), 72.1 (d), 69.9 (t, OCH₂), 35.7 (t), 25.8 (q), 22.7 (t), 20.0 (q), 18.0 (q), 14.2 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.24 (s, 2H, ArH), 4.60 (d, J=7.5 Hz, 2H, OCH₂), 5.49 (t, 1H, CH), 5.06 (m, 1H, CH), 0.87 (t, 3H, CH₃), 1.33 (br t, m, 7H, 2CH₂, 1CH₃), 1.66 (s, 3H, CH₃), 1.72 (s, 3H, CH₃), 6.00 (br, 2H, OH).

Pentan-2-yl 3-(3-methylbut-2-enyloxy)-4,5-dihydroxybenzoate (C₁₇H₂₄O₅, 21): obtained in 22% yield as colorless solid. ¹³C NMR (CDCl₃, 400 MHz) δ : 166.2 (s, CO), 145.6 (s), 143.3 (s), 139.3 (s), 137.0 (s), 122.3 (s), 118.8 (d), 110.7 (d), 106.0 (d), 71.7 (t), 66.0 (t, OCH₂), 35.7 (t), 27.6 (t), 25.6 (q), 22.5 (q), 20.0 (q), 18.2 (q), 14.0 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.29 (s, 1H, ArH), 7.24 (s, 1H, ArH), 4.58 (d, J=6.8 Hz, 2H, OCH₂), 5.08 (m, 1H, OCH), 5.43 (t, 1H, CH), 0.86 (t, 3H, CH₃), 1.71 (s, 3H, CH₃), 1.76 (s, 3H, CH₃), 1.29, 1.27 (br m, 7H, CH₃, 2CH₂).

Pentan-2-yl-4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10-Trienyloxy)-3-hydroxy-5-((2E,6E)-3,7,11-trimethyldodeca-2,6,10-trienyl) benzoate (C42H64O5, 22): obtained in 32% yield as colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 165.9 (s, CO), 151.3 (s), 149.5 (s), 143.9 (s), 141.5 (s), 138.3 (s), 135.5 (s), 135.4 (s), 131.3 (s), 126.2 (s), 114.4 (s), 124.3 (d), 123.6 (d), 119.2 (d), 109.7 (d), 106.6 (d), 71.7 (d), 69.3 (t, OCH₂), 65.7 (t, OCH₂), 39.7 (t), 39.6 (t), 39.6 (t), 39.6 (t), 35.7 (t), 27.6 (t), 26.7 (t), 26.2 (t), 25.9 (t), 22.6 (t), 25.7 (q), 25.7 (q), 20.0 (q), 20.0 (q), 17.7 (q), 17.7 (q), 16.7 (q), 16.4 (q), 16.4 (q), 16.0 (q), 14.0 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.28 (s, 1H, ArH), 7.20 (s, 1H, ArH), 4.69 (d, J=7.4 Hz, 2H, OCH₂), 4.64 (d=6.5 Hz, 2H, OCH₂), (s, 1H, OH), 5.50 (m, 1H, CH), 5.10 (br t, 6H, CH), 0.90 (t, 3H, CH₃), 1.33 (d, J=9.2 Hz, 3H, CH₃), 1.67 (m, 12H, CH₃), 1.71 (m, 12 H, CH₃).

Methyl 3,4,5-tris(methoxymethoxy)benzoate (C₁₄H₂₀O₈, 23): obtained in 85% yield as colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 165.7 (s, CO), 150.2 (s), 150.2 (s), 140.1 (s), 125.3 (s), 110.8 (d), 110.8 (d), 97.9 (t), 94.6 (t), 94.6 (t), 56.5 (q), 55.8 (q), 55.8 (q), 51.6 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 7.54 (s, 2H, ArH), 5.20 (d, J=5.0 Hz, 6H, OCH₂), 3.60 (s, 3H, OCH₃), 3.87 (s, 3H, OCH₃), 3.50 (s, 6H, OCH₃).

1-(3-methylbut-2-enyloxy)-4-methylbenzene (C₁₂H₁₆O, **24):** side product, obtained in 87% yield as colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 156.6 (s, CO), 137.8 (s), 129.8 (d), 129.6 (d), 119.8 (d), 114.3 (d), 114.3 (d), 64.6 (t, OCH₂), 25.7 (q, CH₃), 20.4 (q, CH₃), 18.1 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ : 7.12 (d, 2H, J=8.1 Hz, 2H, ArH), 6.87 (d, J=7.0 Hz, 2H, ArH), 5.57 (t, 1H, CH), 2.35 (s, 3H, CH₃), 1.85 (s, 3H, CH₃), 1.80 (s, 3H, CH₃). The compound is a by-product of the reaction.

4-(3-Methylbut-2-enyloxy)-3-methoxybenzaldehyde

(C13H16O3, 25): side product, obtained in yield 72% as yellow oil. HRESI-MS m/s M+Na⁺ 243.1000, calc. 243.0997; ¹³C NMR (CDCl₃, 400 MHz) δ : 190.7 (s, CHO), 153.7 (s, CO), 149.6 (s, CO), 138.6 (s), 129.7 (s), 126.6 (d), 118.7 (d), 111.4 (d), 108.7 (d), 66.7 (t, OCH₂), 55.8 (q, OCH₃), 25.7 (q, CH₃), 18.1 (q, CH₃); ¹H NMR (CDCl₃, 400 MHz) δ : 9.76 (s, CHO), 7.35 (d, J=8.2 Hz, 1H, ArH), 7.33 (d, J=6.8 Hz, 1H, ArH), 6.90 (d, J=8.1 Hz, 1H, ArH), 5.44 (t, 1H, CH), 4.59 (d, J=6.6 Hz, 2H, CH₂), 3.84 (s, 3H, CH₃), 1.70 (s, 3H, CH₃), 1.68 (s, 3H, CH₃). The compound is the by-product of the reaction.

4-((2E,6E)-3,7,11-trimethyldodeca-2,6,10-trienyloxy)-3methoxybenzaldehyde (C₂₃H₃₂O₃, 26): side product, obtained in 62% yield as yellow colorless oil. ¹³C NMR (CDCl₃, 400 MHz) δ : 191.0 (d, CHO), 153.8 (s), 149.8 (s), 141.7 (s), 135.5 (s), 131.3 (s), 129.9 (s), 126.8 (d), 124.3 (d), 123.5 (d), 118.7 (d), 111.6 (d), 109.7 (d), 66.0 (t, OCH₂), 56.0 (q, OCH₃), 39.7 (t), 39.5 (t), 26.7 (t), 26.1 (t), 24.8 (q), 17.7 (q), 16.8 (q), 16.0 (q); ¹H NMR (CDCl₃, 400 MHz) δ : 9,81 (s, CHO), 7.39 (d, 2H, J= 10.5 Hz, ArH), 6.93 (d, 1H, J=8.1 Hz, ArH), 4.68 (d, 2H, J=8.1 Hz, OCH₂), 3.89 (3H, s, OCH₃), 5.47 (1H, t, CH), 5.05 (2H, t, CH). The compound is the by-product of the reaction.

3,5-dihydroxybenzaldehyde (CrH₆O₃, **27):** obtained in yield 86% as yellow oil. ¹³C NMR (MeOD, 400 MHz) δ : 194.2 (d, CHO), 160.4 (s), 160.4 (s), 140.0 (s), 109.8 (d), 108.7 (d), 108.7 (d); ¹H NMR (MeOD, 400 MHz) δ : 9.75 (s, CHO), 6.78 (br s, 2H, ArH), 6.54 (s, 1H, ArH).

|--|

| Structures of | the prepared | l compounds |
|---------------|--------------|-------------|
|---------------|--------------|-------------|

| No. of the compound | Structure of the compound | |
|---------------------|---------------------------|--|
| 1 | HO OH | |
| 2 | | |
| 3 | | |
| 4 | HO OH OH | |
| 5 | HO OH OH | |
| 6 | HO OH | |
| 7 | HO OH OH | |
| 8 | | |
| 9 | | |
| 10 | | |



Table 3 (continued)

Table 3 (continued)



Biological assay

Mycelial growth inhibition test in vitro

The prepared compounds were dissolved in acetone and tested for antifungal activities *in vitro* by a Poison Food Technique.¹⁴ Potato dextrose agar (PDA) was used as the medium for all test fungi. The media incorporating test compounds at concentration of 50 µg/mL was inoculated at the center of the test fungi in agar discs (4 mm diameter). Three replicate plates for each fungi were incubated at $26\pm2^{\circ}$ C. Control plates containing media mixed with acetone (1 ml) were included. After incubation for 2-6 days, the mycelial growth of fungi (mm) in both treated (T) and control (C) Petri dishes were measured diametrically in three different directions until the fungal growth in the control dishes was almost complete. The percentage of growth inhibition (*I*) was calculated using the formula:

$$I(\%) = (C - T)/C \times 100$$
(1)

The corrected inhibition (IC) was then calculated as follows:

$$IC = (I - CF)/(100 - CF) \times 100$$
 (2)

where $CF = (90 - C_0)/C_0 \times 100$, 90 is the diameter (mm) of the Petri dish, and C_0 is the growth (mm) of the fungus in the control.

Analysis of variance was performed on the data with the PROCGLM procedure (SAS Institute, Cary, NC, USA). If the value of P > F was less than 0.01, means were separated with the least significant different (LSD) test at the p = 0.05 level.

In vivo assay

In order to further investigate the *in vivo* antifungal activities of the synthesized compounds, such as the duration of protection and curative activity, the plant disease of wheat powdery mildew (*Erysiphe graminis*) was used in the test. The effects of the test compounds on disease development and spread were determined using potted plants in a greenhouse.

The potted plants were arranged randomly in two groups in a greenhouse and watered twice daily with tap water. The potted plant seedlings were sprayed with the solutions of test compounds in water/acetone (95:5 v:v) that contained Tween 20 (250 μ g/mL) as wetter, and allowed to stand for 24h.

For the test of preventive effects, the plants in first group were inoculated with the pathogen of the plant disease, one day after being sprayed with either the test compounds or a standard fungicide at dose 500, 250 g/mL. For the test of curative effects, the plants in second group were firstly inoculated with the plant pathogenic fungi, one day before the application of the test compounds and a standard fungicide at dose 500, 250 μ g/mL. Control plants in each group were similarly treated with distilled water/acetone containing Tween 20.

For the development of wheat powdery mildew, the treated wheat seedlings at the first stage were inoculated with *E. graminis* by shaking the infected leaves over them. The inoculated wheat seedlings were incubated for 8 days at $20\pm1^{\circ}$ C and 60% RH (relative humidity) of the day and $18\pm1^{\circ}$ C and 60% RH of the night with 16 h of daylight per day in artificial climate chambers (RP-300, R. P. China), and then the disease severity was determined. The disease severity was recorded on a 0-5 scale, where 0 = no colonies visible to the unaided eye; 1 = few scattered, small discrete colonies; 2 = larger, but still discrete colonies; 3 = colonies merging to form larger mildew lesions; 4 = mildew covering half the total leaf surface and 5 = mildew covering the total leaf surface.¹⁵

The experiment was conducted three times and the mean value of the three estimates for each treatment was converted into percentage fungal control by the equation:

$$\operatorname{control}(\%) = 100 \times (A - B)/A \tag{3}$$

where A = disease incidence (%) on leaves or steam sprayed with Tween 20 solution alone and B = disease incidence (%) on treated leaves or sheaths.

The percentage disease incidence was determined using the formula:

disease incidence (%) = (Σ scale × number of plant leaves infected)/ (highest scale × total number of leaves) × 100 (4)

Analysis of variance was performed on the data with the PROC GLM procedure (SAS Institute, Cary, NC, USA). If the value of P > F less than 0.01, means were separated with the least significant different (LSD) test at the p = 0.05 level.

CONCLUSION

In conclusion, a series of polyphenol ether derivatives were prepared, some of which possess high fungicidal activity. In particular, compound **8**, **13**, **15** and **21** exhibited strong antifungal activities against *R. solani and S. sclerotiorum in vitro*, and compound **13** displayed the best antifungal efficacy toward *E.* graminis *in vivo*. The results suggest that the polyphenol ether derivatives have the potential to be developed as candidates of antifungal products.

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REFERENCES

- E. B. Mojzer, M. K. Hrnčič, M. Škerget, Z. Knez and U. Bren, *Molecules.*, 2016, 21, 901-938.
- Z. Rasines-Perea and P. L. Teissedre, *Molecules.*, 2017, 22, 68-86.
- 3. S. A. Devi and A. Chamoli, *Adv. Exp. Med. Biol.*, **2020**, *1260*, 159-174.
- Y. Zhang, M. Lan, J-P. Lv, J-F. Li, K-Y. Zhang, H. Zhi, H. Zhang and J-M. Sun, *Chem. Biodivers.*, **2020**, *17*, e1900479-e1900489.
- L. Delgado-Roche, K. González, F. Mesta, B. Couder, Z. Tavarez, R. Zavala, I. Hernandez, G. Garrido, I. Rodeiro and W. V. Berghe, *Front. Pharmacol.*, **2020**, *11*, 592985-592995.
- C. Lu, C. Li, B. Chen and Y-H. Shen, *Food Chem.*, 2018, 265, 111-119.
- 7. N. Yahfoufifi, N. Alsadi and M. Jambi, C. Matar, Nutrients., 2018, 10, 1618-1640.
- 8. F. H. A. Fernandes and H. R. N. Salgado, *Crit. Rev. Anal. Chem.*, **2016**, *46*, 257-265.
- D. Wang and B. Bao, Drug. Des. Devel. Ther., 2020, 14, 1583-1592.
- N. M. Aborehab and N. Osama, *Cancer Cell Int.*, 2019, 19, 154-166.
- C. L. Tsai, Y. M. Chiu, T. Y. Ho, C. T. Hsieh, D. C. Shieh, Y. J. Lee, G. J. Tsay and Y. Y. Wu, *Anticancer Res.*, **2018**, 38, 2057-2067.
- Y. L. Liu, C. C. Hsu, H. J. Huang, C. J. Chang, S. H. Sun and A. M. Y. Lin, *Mol. Neurobiol.*, **2020**, *57*, 96-104.
- D. Karas, J. Ulrichová and K. Valentová, Food Chem. Toxicol., 2017, 105, 223-240.
- M. Agarwal, S. Walia, S. Dhingra and B. P. S. Khambay, *Pest Manag. Sci.*, 2001, 57, 289–300.
- K. D. Hickey, "Methods for evaluating pesticides for control of plant diseases", **1986**, p. 103–108.